Porcine Gastrointestinal Pathology and Laboratory Diagnosis of Disease

Eric R. Burrough, DVM, PhD, DACVP

Professor, Diagnostic Pathologist, Pathology Section Leader Iowa State University Veterinary Diagnostic Laboratory



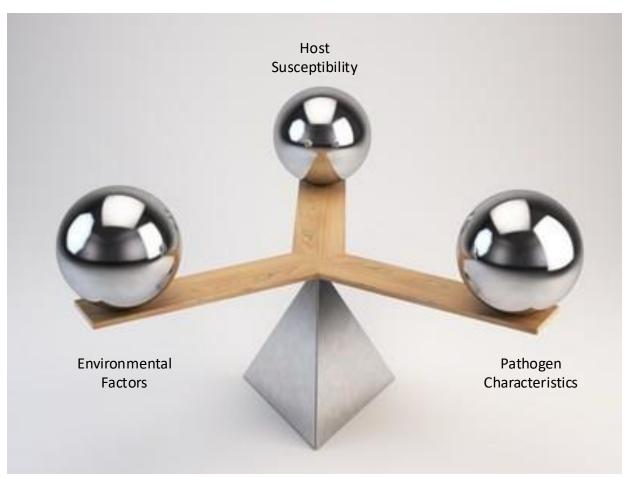
- Goals for today's talk:
 - Present common infectious diseases of pigs
 - Diseases that are particularly relevant for Australian pigs
 - Utilize case material from recent diagnostic cases in the US
 - Over 90% of the materials used are from personal cases received since 2023

Veterinary Diagnostic

- Demonstrate commonly used ancillary diagnostic tools in the US
- Provide visual examples of common lesions
 - Quality examples of classic lesions
 - Align gross features with histologic lesions
 - Review common histology terms in pathology reports
 - What phrases suggest different disease processes?



- Health challenges in commercial pigs can be a diagnostic dilemma:
 - Many common enteric pathogens are endemic on affected farms
 - Detection may or may not = disease







- Health challenges in commercial pigs can be a diagnostic dilemma:
 - Many common enteric pathogens are endemic on affected farms
 - Detection may or may not = disease
 - Available diagnostic tests may not differentiate pathogens / non-pathogens / vaccines
 - PCR, culture, ELISA, even IHC
 - MLV vaccines are common in swine production (PRRSV, Lawsonia, Salmonella)
 - **Disease expression is variable** within and among farms
 - On-farm management factors impact disease expression
 - Mixed infections are common (if not the norm)
 - The key to diagnostic accuracy is the **right sample** from the **right animal** at the **right time**

Veterinary Diagnostic



- Health challenges in commercial pigs can be a diagnostic dilemma:
 - What is wrong with this diagnostic submission for enteric disease?





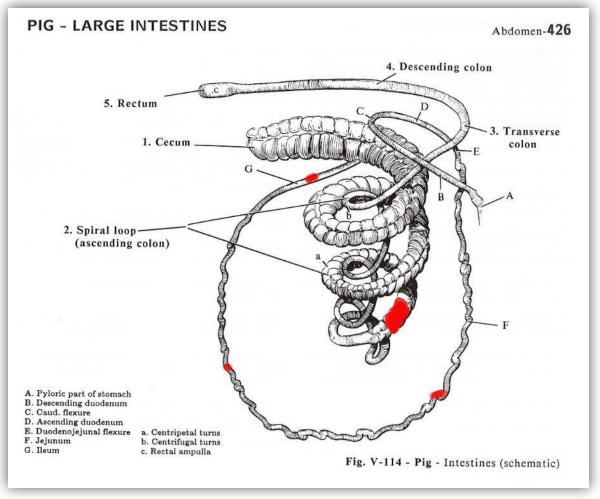
- Health challenges in commercial pigs can be a diagnostic dilemma:
 - What is wrong with this diagnostic submission for enteric disease?
 - Looks SOP driven:
 - Sacrifice 3 pigs
 - Collect fresh and fixed samples of all major internal 'clean' organs
 - Collect fresh and fixed small and large intestine
 - 2 3 sections (approx. 1") of small intestine in formalin; the rest fresh
 - 1 small random section of colon fixed; the rest fresh
 - Put it all in a box and send to the diagnostic laboratory
 - All major systems were sampled; however, if the diagnostic question is causes of enteric disease, why not focus more on the intestines?

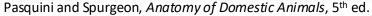
Veterinary Diagnostic

- Histopathology has high diagnostic specificity but relatively poor diagnostic sensitivity
- There are many feet of intestine and only 3 inches were fixed for evaluation
 - Enteric diseases are often multifocal or segmental in nature



- Health challenges in commercial pigs can be a diagnostic dilemma:
 - What is wrong with this diagnostic submission for enteric disease?







- Health challenges in commercial pigs can be a diagnostic dilemma:
 - How can we improve this submission for enteric disease?
 - Refine the SOP for enteric disease:
 - Sacrifice 2-3 pigs with active clinical disease consistent with the issue
 - Collect fresh and fixed samples of all major internal 'clean' organs
 - Collect fresh and fixed samples of internal 'clean' organs that look abnormal
 - Collect fresh and fixed small and large intestine
 - **5 6 sections** (approx. 1") of intestine in formalin, **especially any that look abnormal**; some fresh
 - 2 3 sections of colon fixed, especially abnormal regions; some fresh (can separate some content)
 - Put it all in a box and send to the VDL
 - Different diseases affect different segments:
 - Whipworms are commonly observed in the cecum
 - Dysentery is most often found in the spiral colon (apex)
 - Salmonella lesions are more commonly observed in the centripetal spiral than SI
 - Enteric viruses commonly impact the aboral jejunum and ileum more then other segments

- Coccidia are commonly observed in the ileum
- Lawsonia lesions can occur from jejunum through the spiral colon

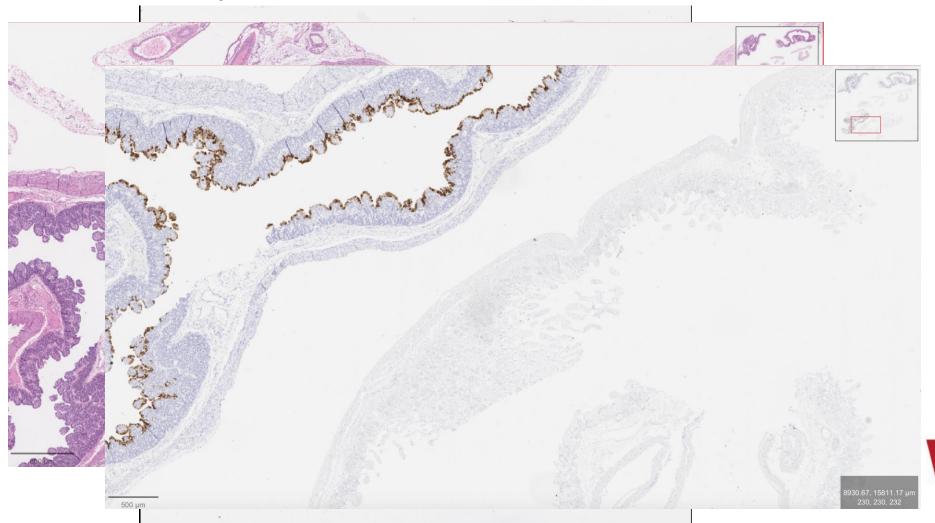


- Health challenges in commercial pigs can be a diagnostic dilemma:
 - What about autolyzed intestinal tissue?
 - PCR
 - Minimal to no impact on direct sample PCR
 - Would not be ideal for NGS or microbiome evaluation
 - Culture
 - In most cases is still okay,
 - Will have some overgrowth of some bacteria, may lose fastidious organisms
 - Histopathology
 - Can often still see villus length and assess villus:crypt ratio
 - Typically lose the mucosa:
 - No way to confirm ETEC or coccidia
 - IHC for viral agents may be falsely negative





- Health challenges in commercial pigs can be a diagnostic dilemma:
 - What about autolyzed intestinal tissue?



Common endemic infectious disea	ises in US swine		
Etiologic agent	Age often affected	Clinical signs	Microscopic lesions
Enteric pathogens			
Coronaviruses	All ages	Acute watery diarrhea, can be severe in neonates	Segmental villus blunting and fusion that can be severe; epithelial necrosis; mild inflammation
Clostridium perfringens type C	Birth to 7 days	Hemorrhagic diarrhea; rapid death often of an entire litter	Mucosal necrosis, suppuration, hemorrhage, emphysema; gram- positive rods; diphtheritic membrane
Clostridioides difficile	Birth to 2 weeks	Creamy diarrhea; dehydration; usually mild but can be severe if poor immunity	Erosive typhlocolitis with multifocal "volcano" lesions, suppuration, mesocolonic edema
Rotavirus	Birth to 6 weeks	Creamy diarrhea; dehydration; usually mild; decreased growth rate	Segmental villus blunting; epithelial necrosis; mild inflammation
E. coli	Birth to 7 weeks	Acute diarrhea, watery, severe; dehydration; neonates (1-7 days) or post-weaning (3-7 weeks)	None; short bacterial rods attached to enterocytes; congestion
Coccidia	1 - 4 weeks	Creamy diarrhea; dehydration; usually mild; decreased growth rate	Villous atrophy, fibrinonecrotic enteritis, intracellular merozoites
Salmonella	After 3 weeks	Diarrhea, mild to severe, may see fibrin flecks; fever	Ulceration and suppuration; colon often more severely affected
Lawsonia	After 5 weeks	Diarrhea, intermittent or severe with blood, pallor, rapid death; decreased growth	Crypt enterocyte hyperplasia, branching crypts, loss of goblet cells; necrosis; curved rods within epithelial cells
Brachyspira spp.	After 5 weeks	Mucoid diarrhea, +/- blood (dysentery); dehydration; reduced gain; deaths	Erosion of epithelium; goblet-cell hyperplasia, mixed inflammatory infiltrates, fibrinous exudate





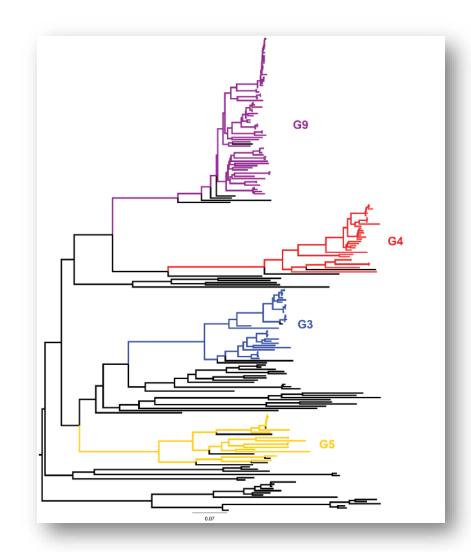
- Small Intestinal Diseases
 - Enteric viruses
 - Coronaviruses -> rotaviruses -> sapovirus
 - Many farms are endemically infected
 - Disease expression varies by immune status
 - Rotaviruses have a segmented genome and reassortment is common
 - Immunity not cross-protective among RVs or Cvs
 - Mixed infections are common
 - Gross lesions are all similar
 - Thin-walled small intestine
 - Watery contents
 - Reduced/absent chyle in lymphatics







- Small Intestinal Diseases
 - Enteric viruses
 - Coronaviruses -> rotaviruses -> sapovirus
 - Diagnostics
 - Histopathology
 - Segmental disease, may need to look at multiple segments
 - PCR
 - Everything flows downstream
 - Cecal/colon content is best
 - Sequencing
 - Predict genotype/serotype
 - Match vaccines
 - Develop custom RNA vaccines
 - Direct detection (IHC, ISH)



RVA VP7 Sequences at ISU VDL in 2020





- Small Intestinal Diseases
 - Enteric viruses
 - CoVs -> rotaviruses -> sapovirus



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Veterinary

Diagnostic Laboratory



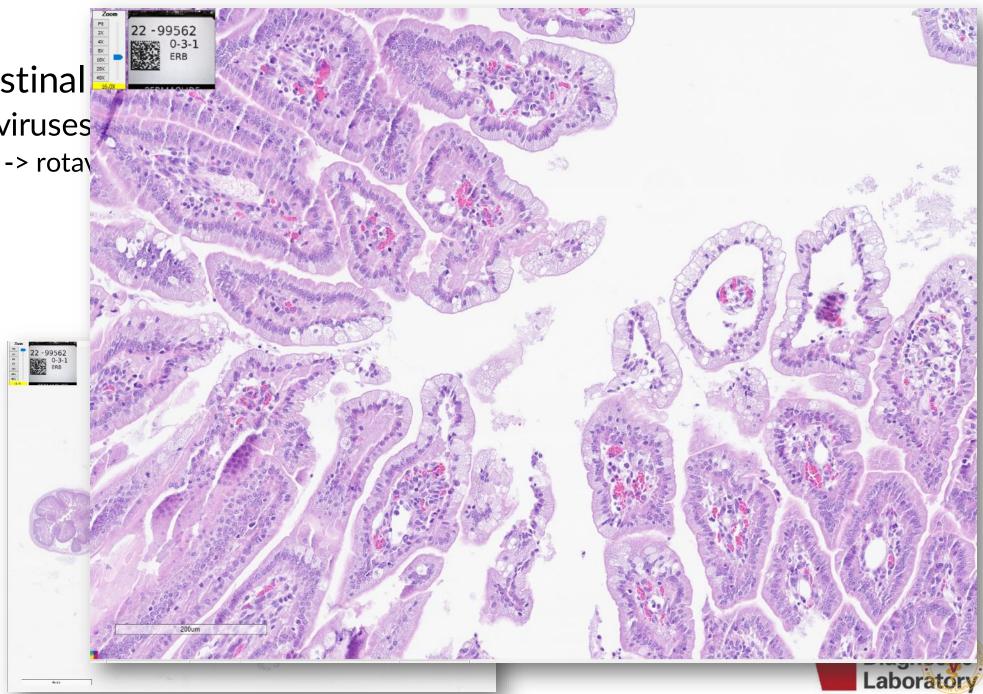
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Small Intestinal

- Enteric viruses
 - CoVs -> rotav

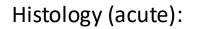
Histology (acute):

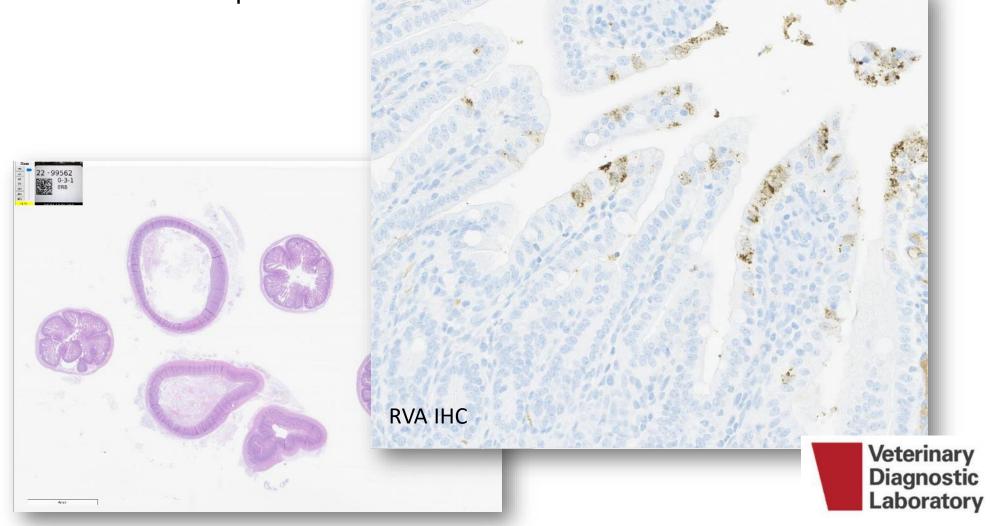
- Vacuolation of villus tip enterocytes
- Possible syncytia





- Small Intestinal Diseases
 - Enteric viruses
 - CoVs -> rotaviruses -> sapovirus





- Small Intestinal Diseases
 - Enteric viruses
 - CoVs -> rotaviruses -> sapovirus

Histology (subacute):

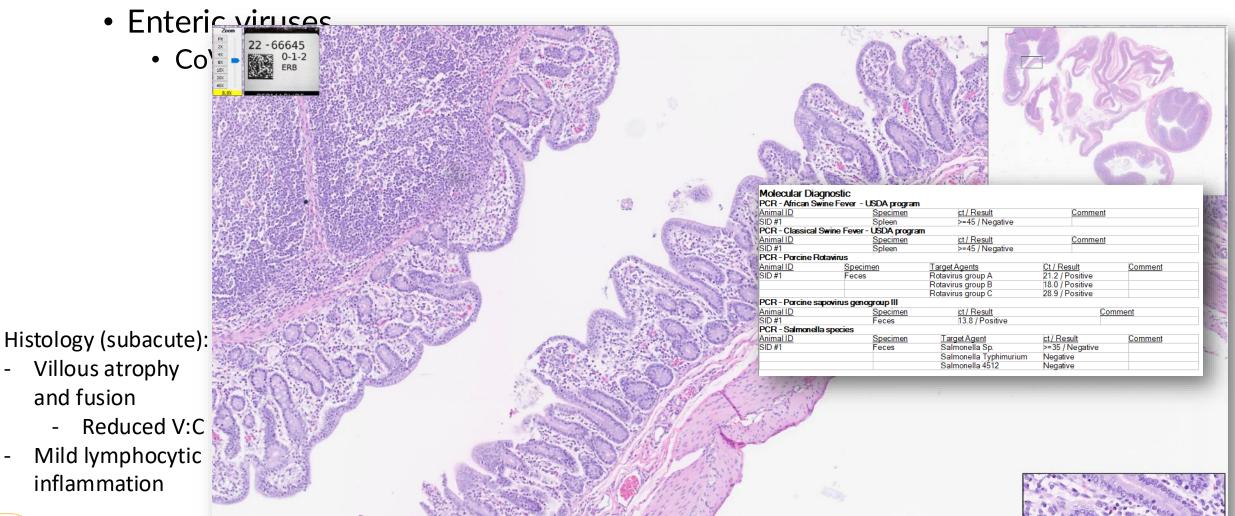
- Villous atrophy and fusion
 - Reduced V:C -
- Mild lymphocytic inflammation





• Small Intestinal Diseases

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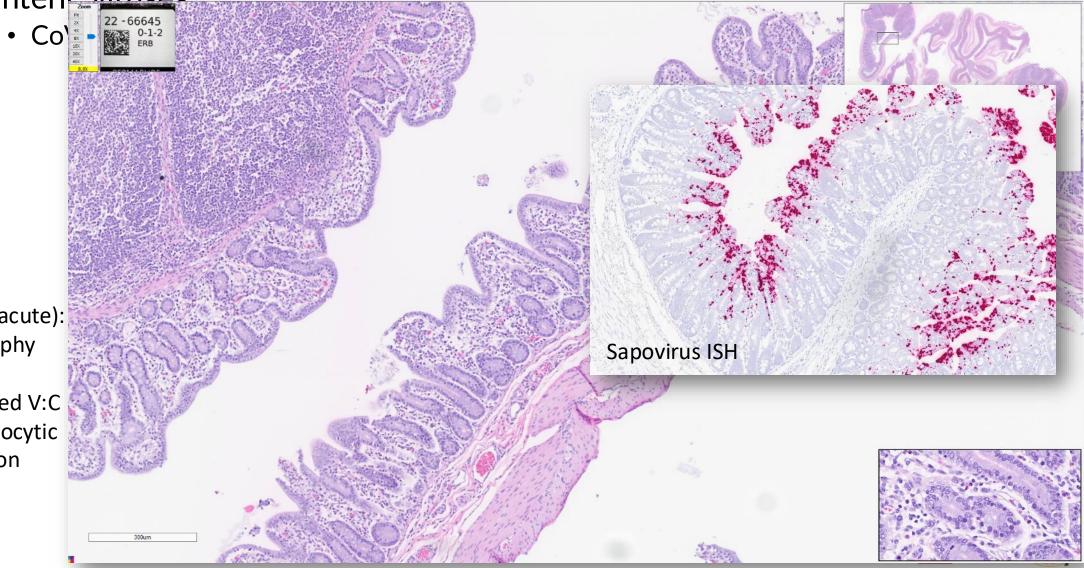
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- Small Intestinal Diseases
 - Enteric viruses

- Histology (subacute):
- Villous atrophy and fusion
 - Reduced V:C
- Mild lymphocytic inflammation



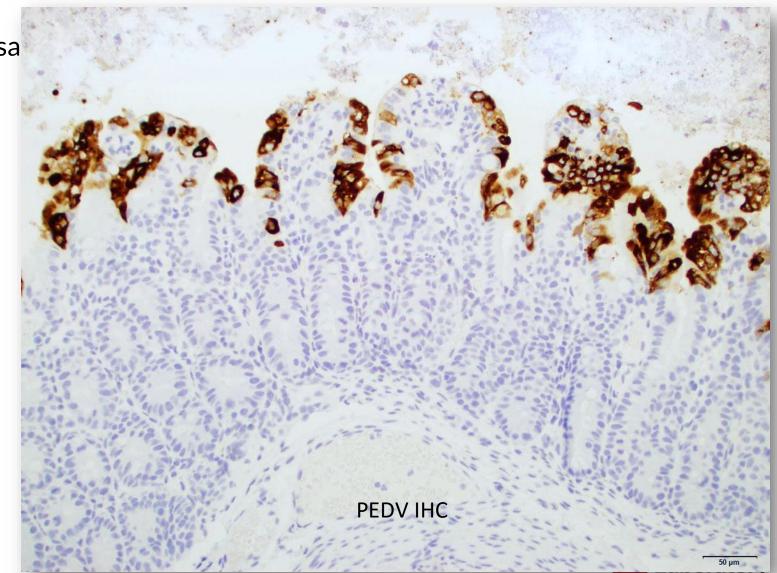


- Small Intestinal Diseases
 - Enteric viruses
 - CoVs -> rotaviruses -> sa

Histology (subacute):

- Villous atrophy and fusion
 - Reduced V:C
- Mild lymphocytic inflammation





- Small Intestinal Diseases
 - Clostridium perfringens Type A
 - Non-specific diarrhea in neonates
 - Low mortality
 - Disease is not consistently reproducible
 - Diagnostics
 - No defined diagnostic criteria
 - No consistent histologic lesions
 - Diagnosis by exclusion
 - Culture of limited use
 - Part of normal flora
 - Poor diagnostic specificity

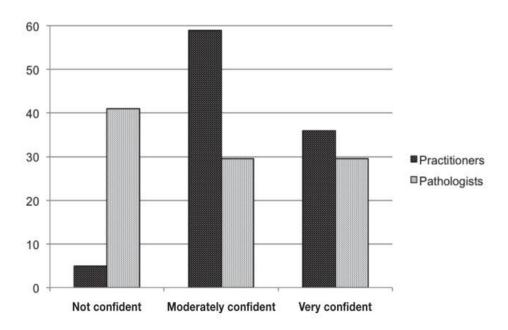






- Small Intestinal Diseases
 - Clostridium perfringens Type A

How do swine practitioners and veterinary pathologists arrive at a diagnosis of *Clostridium perfringens* type A enteritis in neonatal piglets?

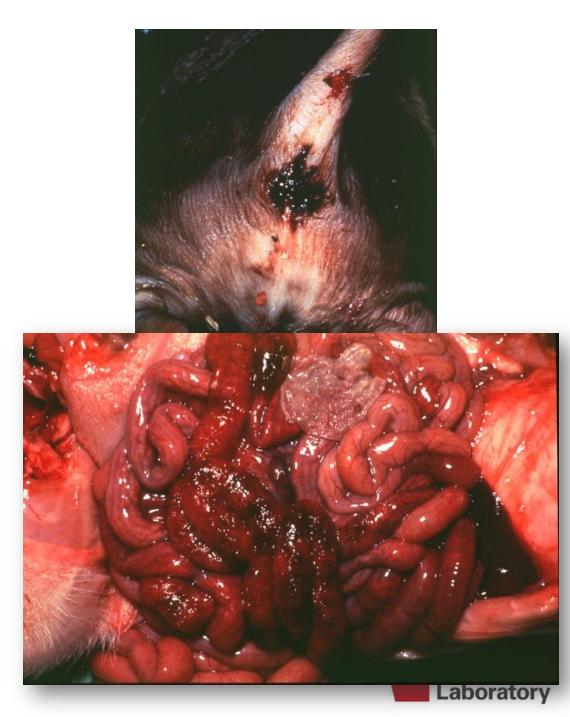


Over 90% of swine practitioners are moderately to very confident diagnosing *C. perfringens* type in A disease, while over 70% of veterinary pathologists are not confident to moderately confident making the same diagnosis!



Chan G, Farzan A, Prescott J, Friendship R. 2013. Can Vet J 54(5):504-506

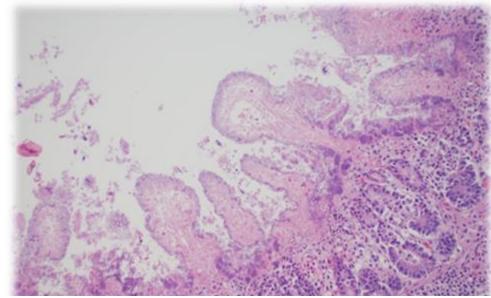
- Small Intestinal Diseases
 - Clostridium perfringens Type C
 - Bloody diarrhea in neonates
 - 3 days of age is common
 - Disease due to beta toxin
 - trypsin-labile
 - High mortality with short course
 - Not all litters are affected
 - Death of entire litter may occur
 - Gross lesions:
 - Segmental hemorrhagic enteritis
 - May have emphysema
 - Perforation and peritonitis possible





- Small Intestinal Diseases
 - Clostridium perfringens Type C
 - Diagnostics:
 - Histopathology
 - Necrosis of luminal portion of villi
 - Lined by bacilli
 - ELISA for direct toxin detection
 - Routine Culture
 - Best if from intestine

• Need genotyping of isolate



Bacteriology

Animal ID	Specimen	Enrichment	Growth	Organism	Comment
o27649, SID #1	Colon		High	Smooth/mucoid	
			-	Escherichia coli	
o27649, SID #1	Colon		High	Clostridium perfringens	
o27649, SID #1	Intestine		High	Smooth/mucoid	
				Escherichia coli	
o27649, SID #1	Intestine		High	Clostridium perfringens	
o27649, SID #1	Lung			No Significant Growth	
o27649, SID #1	Spleen			No Significant Growth	
o36848, SID #2	Colon		High	Clostridium perfringens	
o36848, SID #2	Colon		Heavy	Smooth/mucoid	
				Escherichia coli	
o36848, SID #2	Intestine		High	Clostridium perfringens	
o36848, SID #2	Lung		Few	Streptococcus suis	
				serotype 5	
o36848, SID #2	Spleen			No Growth	

Molecular Diagnostic

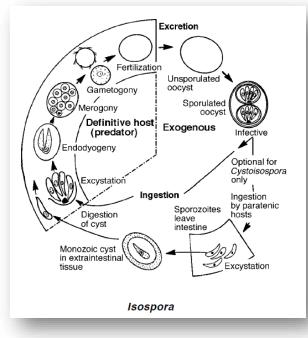
Clostridium perfringens Genotyping Animal ID beta2 Genotype epsilon enterotoxi iota . SID #1 Positive Positive Positive Negative Negative Negative Comment Positive 2. SID #2 Positive Negative Negative Negative Negative Α Comment Diagnostic

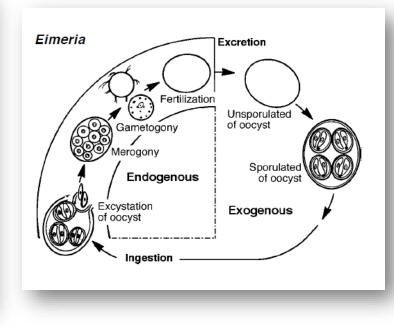
Laboratory



- Small Intestinal Diseases
 - Coccidia
 - Cystoisospora suis (also Eimeria spp.)
 - Most often observed in summer
 - Diarrhea in pre-weaned and recently weaned pigs
 - Inconsistent oocyst shedding
 - Fecal flotation often low yield

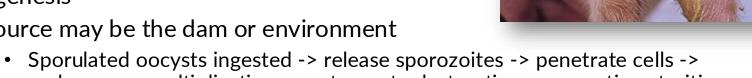








- Small Intestinal Diseases
 - Coccidia
 - Cystoisospora suis (also Eimeria spp.)
 - Most often observed in summer
 - Diarrhea in pre-weaned and recently weaned pigs
 - Inconsistent oocyst shedding
 - Fecal flotation often low yield
 - Pathogenesis
 - Source may be the dam or environment



- endogenous multiplication -> enterocyte destruction -> necrotic enteritis
- Malabsorptive diarrhea (creamy to pasty) ensues
- Affected animals often have lower body condition
- Mortality rates may be low, but increase susceptibility to other enteric agents
- Seasonality may be observed (warmer months)
 - C. suis oocysts sporulate within 24 hours in warm, moist environment
 - Eimeria take longer (~5 days)



Veterinary

Diagnostic

Laboratory



- Small Intestinal Diseases
 - Coccidia
 - Cystoisospora suis (also Eimeria spp.)
 - Most often observed in summer
 - Diarrhea in pre-weaned and recently weaned pigs
 - Inconsistent oocyst shedding
 - Fecal flotation often low yield
 - Fibrinonecrotic enteritis grossly in later stages (secondary)
 - Diagnostics
 - Microscopic lesions of villous atrophy +/- exudate
 - Intraepithelial coccidia
 - Merogony more common with Cystoisospora
 - Gametogony more common with Eimeria
 - Fecal flotation of limited sensitivity
 - PCR available, limited specificity



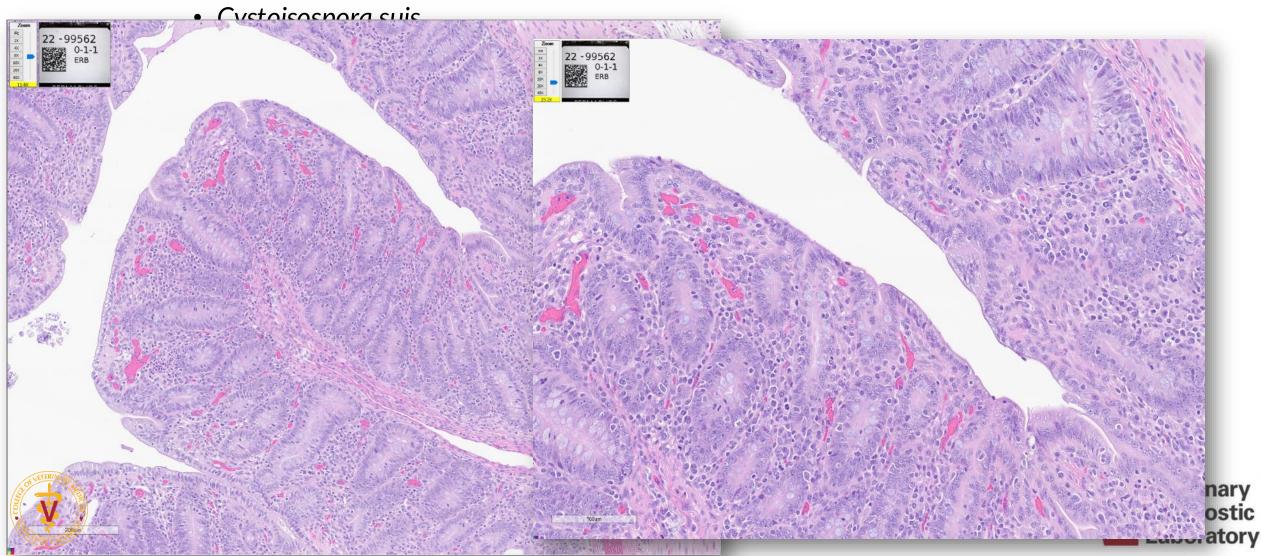


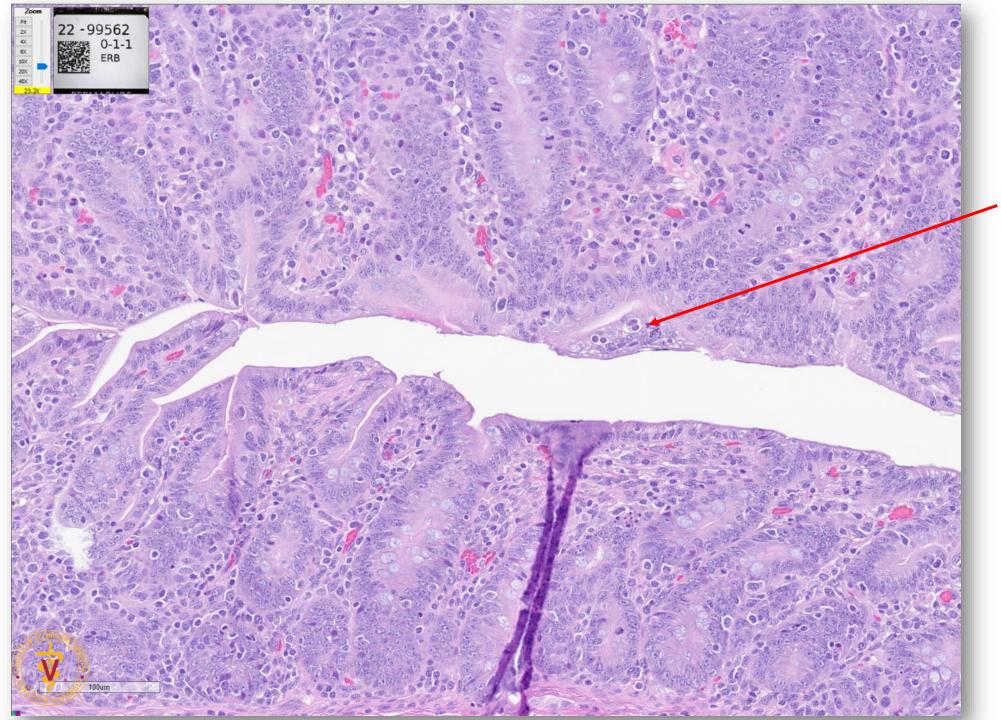


- Small Intestinal Diseases
 - Coccidia
 - Cystoisospora suis



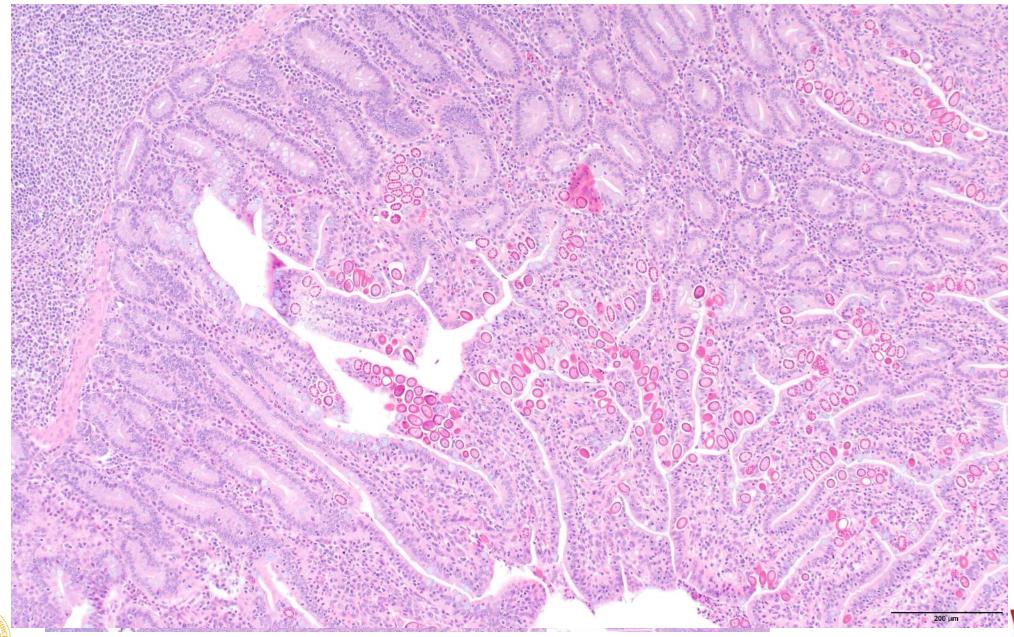
- Small Intestinal Diseases
 - Coccidia





Intraepithelial meronts containing paired type I merozoites typical of *Cystoisospora suis*







- Small Intestinal Diseases
 - Colibacillosis
 - Hemolytic E. coli
 - Disease is commonly observed immediately post-weaning
 - Watery diarrhea, dehydration
 - Some unexpected death
 - Gross lesions are very characteristic
 - This is a secretory diarrhea
 - pH is alkaline (≥8.0)
 - Some ETEC strains harbor Stx2e gene
 - May manifest in edema disease



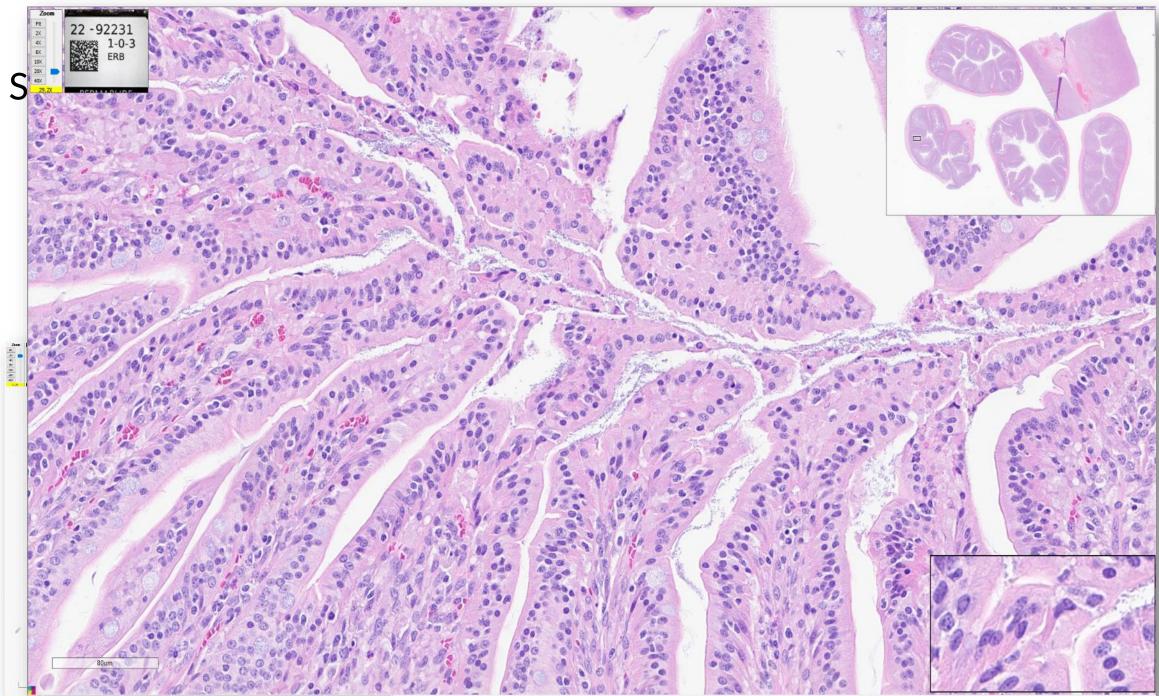


- Small Intestinal Diseases
 - Colibacillosis
 - Diagnostics:
 - Routine culture:
 - In pigs, not all hemolytic *E. coli* are pathogens, but most pathogenic *E. coli* are hemolytic
 - Suckling piglets may be affected by:
 - K88 (F4) hemolytic
 - K99 (F5), F41, 987P non-hemolytic
 - Weaned pigs are affected by:
 - K88 and F18 both hemolytic
 - Hemolysis is a phenotypic trait
 - Colonies can be rough, intermediate, smooth, smooth/mucoid, or mucoid types
 - Genotyping
 - Performed on isolated colonies (can also be done of scrolls from FFPE tissues)

- Must possess both fimbria and enterotoxin genes to be ETEC
- Serotyping is not commonly performed in swine diagnostics (e.g. O157:H7 STEC)
 - Some research papers discuss serovirotype

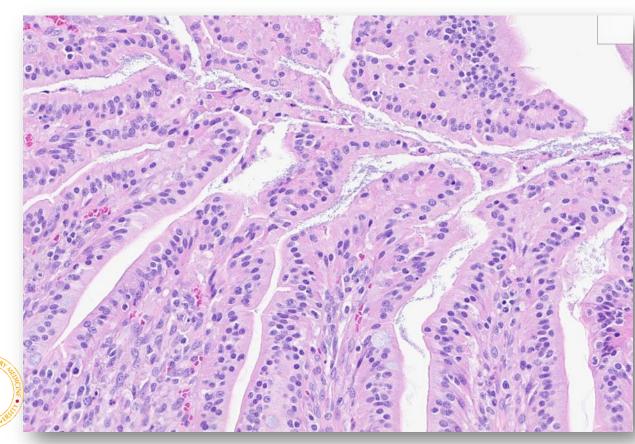






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- Small Intestinal Diseases
 - Colibacillosis
 - Hemolytic E. coli



Bacteriology

Culture Sun	nmary				
Animal ID	Speci	men Enrichment	Growth	Organism	Comment
GA [1-3]	Colon		Single	Salmonella species group B	
GA [1-3]	Intesti	ne	Heavy	Ēscherichia coli - hemolytic (smooth/mucoid)	
Food Anima	al Susceptibility				
GA [1-3]	Final Result	S.SB> Salmonella group B	aspecies		
GA [1-3]	Final Result	E.COLH> Escher haemolytic	ichia coli		

Antimicrobial	S.SB	ECOLH	
	*Int/MIC	*Int/MIC	
Ampicillin	R />16.0000	R />16.0000	
Ceftiofur	S/1.0000	S/0.5000	
Clindamycin	NI / >16.0000	R />16.0000	
Danofloxacin	NI/<=0.1200	NI / >1.0000	
Enrofloxacin	S/<=0.1200	R />2.0000	
Florfenicol	/ 4.0000	R />8.0000	
Gamithromycin	NI / 8.0000	NI / 4.0000	
Gentamicin	S/<=1.0000	1/8.0000	
Neomycin	S/<=4.0000	R/16.0000	
Penicillin	R />8.0000	R />8.0000	
Sulfadimethoxine	R />256.0000	R />256.0000	
Spectinomycin	S/32.0000	R />64.0000	
Tetracycline	R />8.0000	R />8.0000	
Tiamulín	R />32.0000	R />32.0000	
Tildipirosin	NI / 8.0000	NI / 4.0000	
Tilmicosin	R />16.0000	R />16.0000	
Trimethoprim/Sulphamethoxazole	S/<=2.0000	R />2.0000	
Tulathromycin	NI/<=8.0000	NI / <=8.0000	
Tylosin (Tartrate/Base)	NI/>32.0000	NI/>32.0000	
Positive Growth Control	OK/3610.3333	OK / 3512.6667	

Molecular Diagnostic

Animal ID	Gene	Result	
GA [1-3]	EAST1 (toxin)	Positive	
	LT(toxin)	Positive	
	STa(toxin)	Positive	
	STb(toxin)	Positive	
	Stx1 (toxin)	Negative	
	Stx2 (toxin)	Positive	
	Stx2e(toxin)	Positive	
	F18(pilus)	Positive	
	F41(pilus)	Negative	
	K88(pilus)	Negative	
	K99(pilus)	Negative	
	987P(pilus)	Negative	
	AIDA (adhesin)	Negative	
	EAEA (adhesin)	Negative	
	PAA (adhesin)	Negative	
	Organism	Escherichia coli haemolytic	



- Panintestinal Diseases
 - Salmonellosis
 - Serovars 4,[5],12:i:-, Typhimurium, Choleraesuis are known enteropathogens
 - All serovars are potential opportunists \rightarrow commonly follow viral enteritis

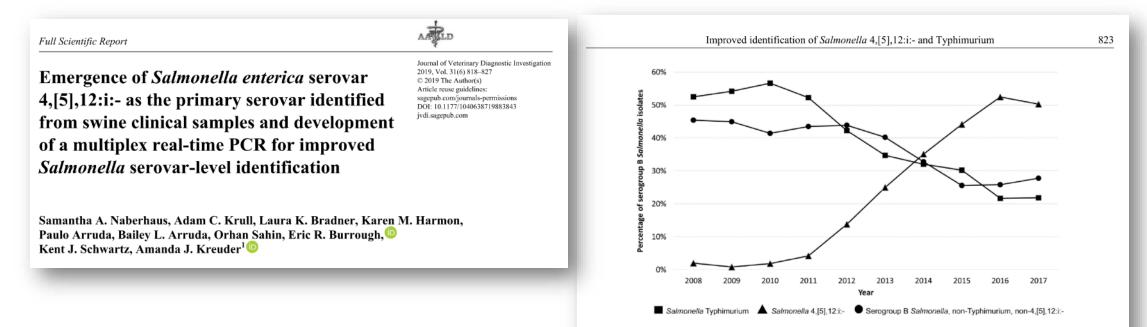


Figure 1. The percent contribution of Salmonella Typhimurium and 4,[5],12:i:- to the serogroup B isolates of Salmonella identified from swine clinical cases at the Iowa State Veterinary Diagnostic Laboratory from 2008 to 2017.





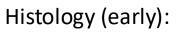
- Panintestinal Diseases
 - Salmonellosis
 - Serovars 4,[5],12:i:-, Typhimurium, Choleraesuis
 - Gross lesions are multifocal to diffuse ulcerative enterocolitis
 - Often pseudomembrane formation and enlarged mesenteric lymph nodes
 - Diagnostics
 - Histopathology
 - Routine culture +/- enrichment
 - Followed by serotyping
 - PCR





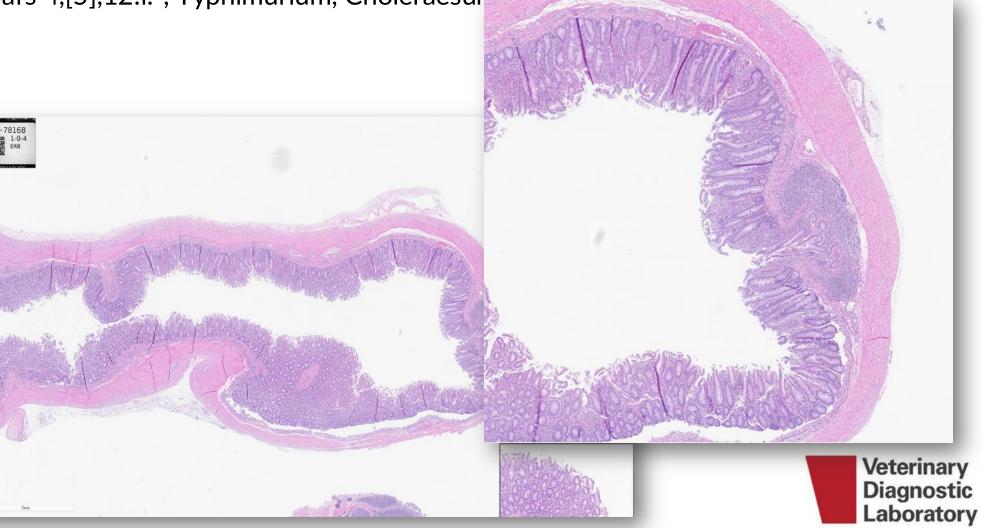


- Panintestinal Diseases
 - Salmonellosis
 - Serovars 4,[5],12:i:-, Typhimurium, Choleraesui



- Shallow erosions
- Scant proprial inflammation
- Neutrophils in crypts



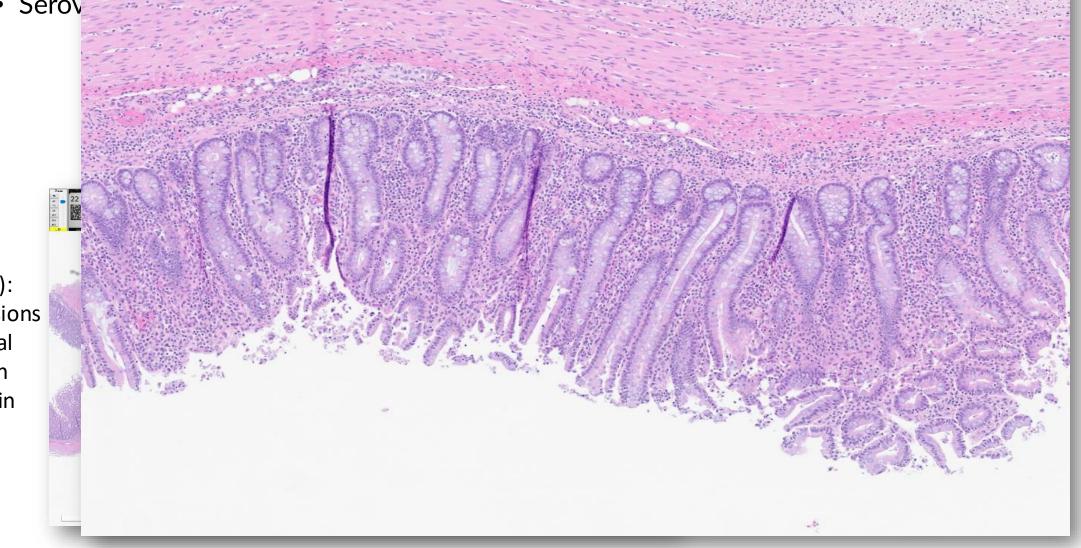


- Panintesti
 - Salmone
 - Serov

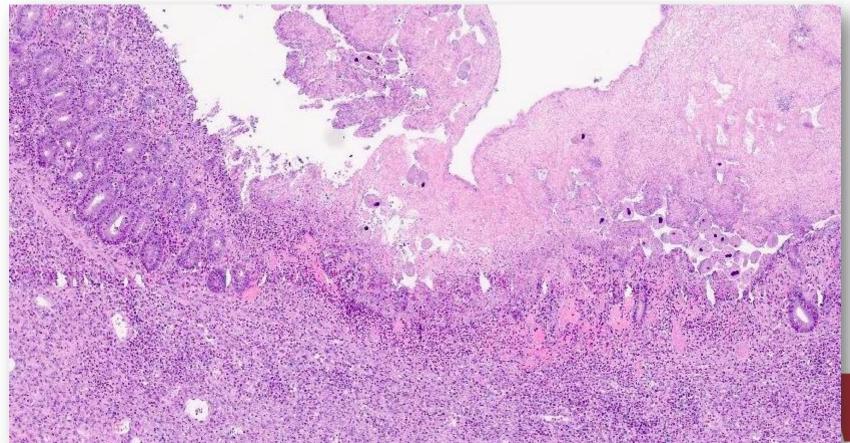
Histology (early):

- Shallow erosions -
- Scant proprial inflammation
- Neutrophils in _ crypts





- Panintestinal Diseases
 - Salmonellosis
 - Serovars 4,[5],12:i:-, Typhimurium, Choleraesuis



Histology (late):

- Deep ulcerations
- Fibrosis
- Inflammation into submucosa
- Colonization by ciliates

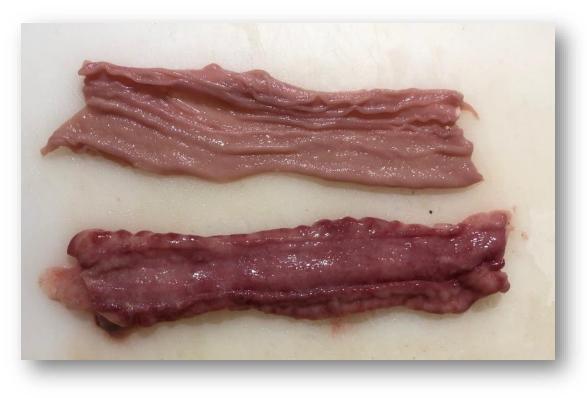


- Panintestinal Diseases
 - Lawsonia intracellularis
 - Obligate intracellular bacterium
 - Grows freely in the cytosol, passes to each daughter cell in mitosis
 - Infected cells do not mature \rightarrow immortalized
 - Hence the original disease name "porcine intestinal adenomatosis"
 - Endemic in many pig populations
 - Disease most often in grow-finish age as immunity wanes
 - Subclinical disease is common
 - Three clinical forms
 - Porcine intestinal adenomatosis (PIA)
 - Necrotic enteritis (NE)
 - Proliferative hemorrhagic enteropathy (PHE)





- Panintestinal Diseases
 - Lawsonia intracellularis
 - Gross lesions
 - PIA form
 - Typically reported in the ileum
 - Can be anywhere from jejunum through spiral colon
 - Thickened, crowded mucosal folds adjacent to Peyer's patches
 - Often superficial congestion



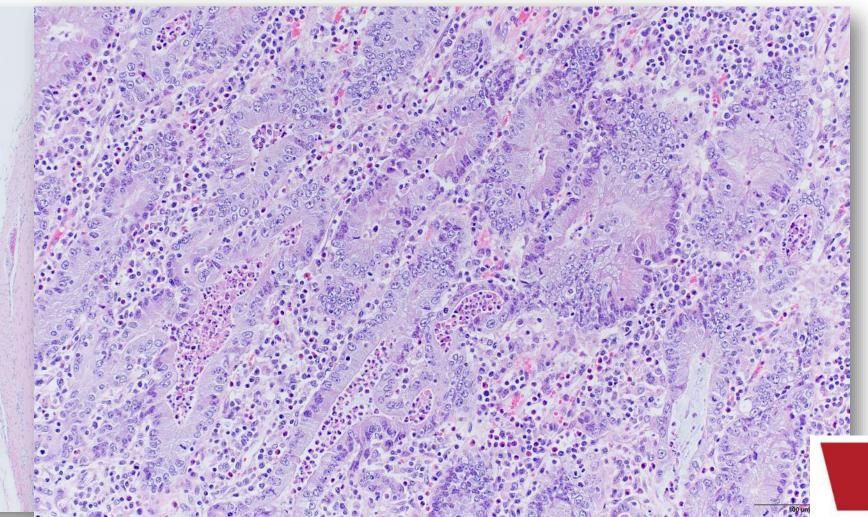




- Panintestinal Diseases
 - Lawsonia intracellularis

Histology (PIA):

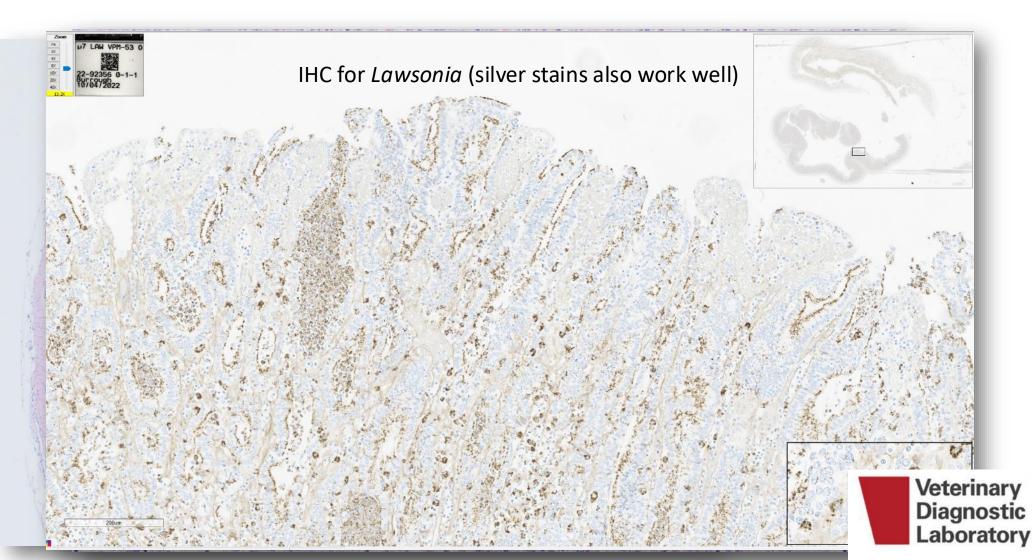
- Swollen
 epithelial cells
 with abundant
 apical
 cytoplasm
- Reduced goblet cells
- Increased mitoses
- Neutrophils in crypts



- Panintestinal Diseases
 - Lawsonia intracellularis

Histology (PIA):

- Swollen
 epithelial cells
 with abundant
 apical
 cytoplasm
- Reduced goblet cells
- Increased mitoses
- Neutrophils in crypts





- Panintestinal Diseases
 - Lawsonia intracellularis
 - Gross lesions
 - NE form
 - Occurs in PIA form with chronicity and secondary bacterial involvement
 - Often marked pseudomembrane formation

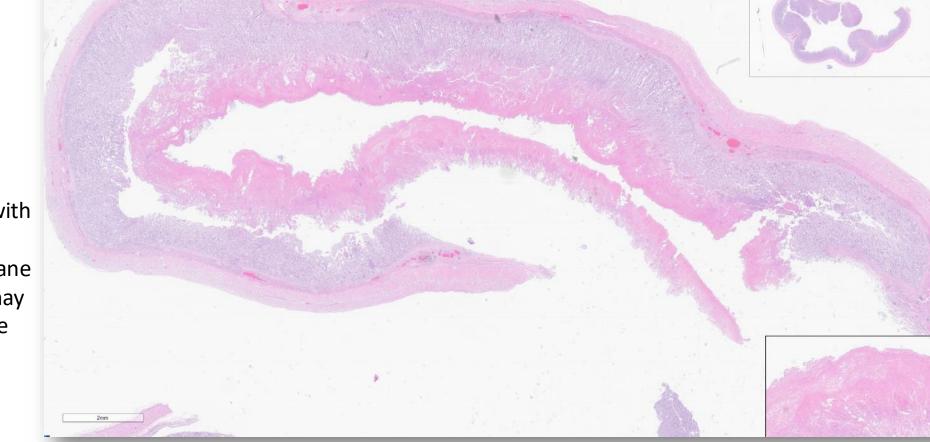






- Panintestinal Diseases
 - Lawsonia intracellularis

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Veterinary

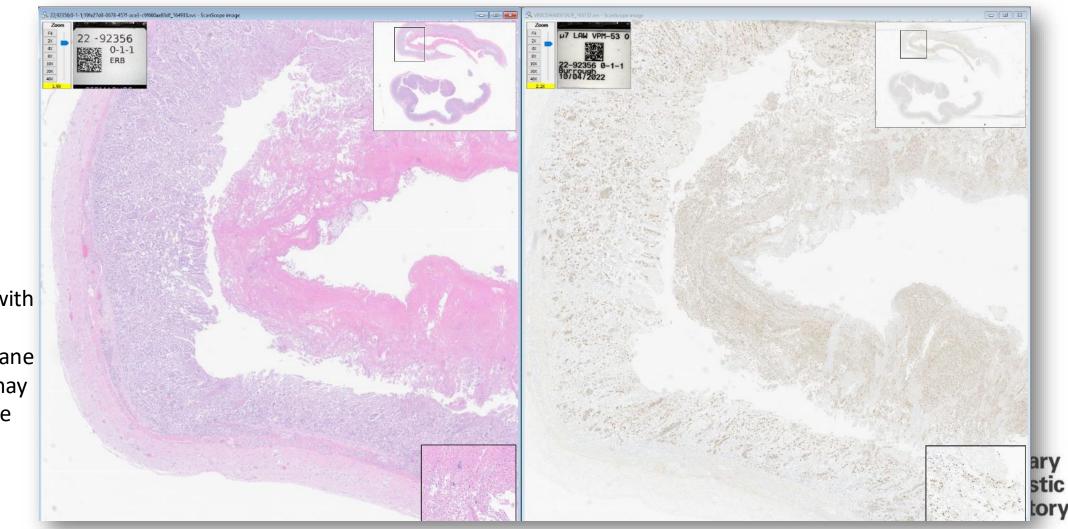
Diagnostic Laboratory

Histology (NE):

- Similar to PIA with added pseudomembrane
- Crypt lesions may begin to resolve



- Panintestinal Diseases
 - Lawsonia intracellularis



Histology (NE):

- Similar to PIA with added pseudomembrane
- Crypt lesions may begin to resolve



- Panintestinal Diseases
 - Lawsonia intracellularis
 - Gross lesions
 - PHE form
 - The most acute form observed
 - Manifests as unexpected death
 - Commonly in young breeding-age gilts
 - Ileal lumen contains clotted blood and may contain a cast of fibrin-enmeshed debris
 - Microscopic features are similar but less severe than PIA form
 - Diagnostics
 - PCR
 - Feces, oral fluids
 - Histopathology + IHC or silver stains
 - Serology (herd monitoring)







- Large Intestinal Diseases
 - Clostridioides difficile
 - Most sows are colonized
 - Piglets typically acquire CD in first 48 hours of life
 - Disease typically occurs in first week of life
 - Toxin-medicated disease
 - Precise triggers unknown
 - Pasty to watery diarrhea
 - Induces mesocolonic edema
 - Diagnostics
 - Histopathology
 - Toxin ELISA (TcdA, TcdB)?



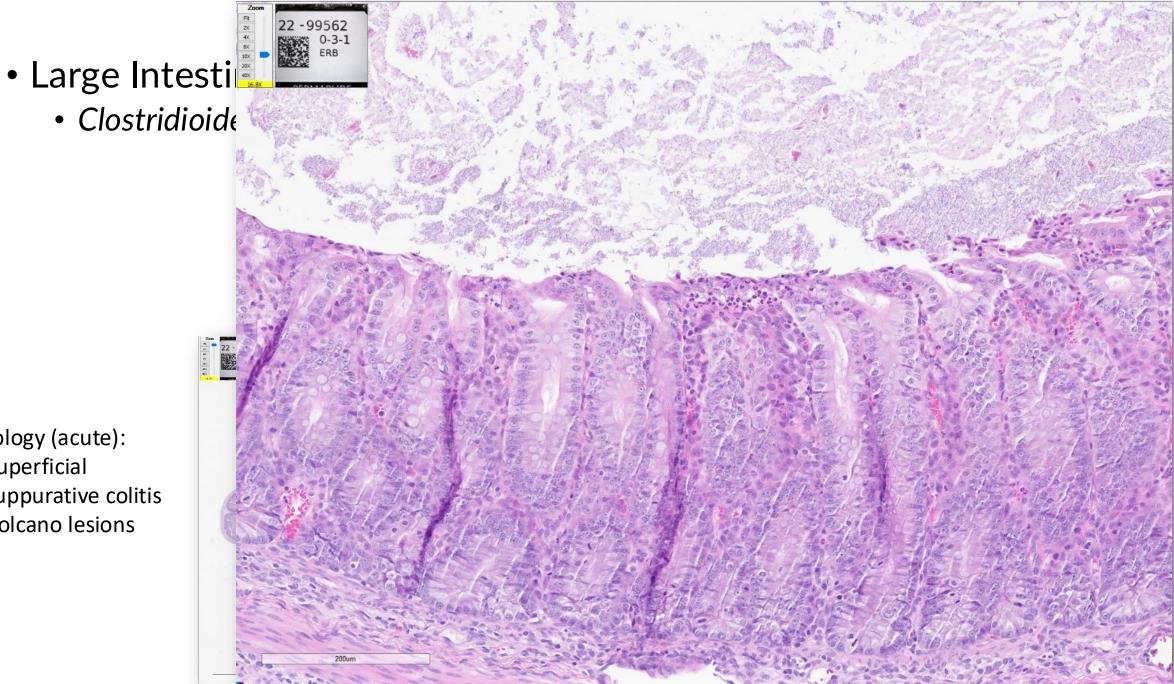


- Large Intestinal Diseases
 - Clostridioides difficile



Histology (acute):





Histology (acute):

- Superficial suppurative colitis
- Volcano lesions

- Large Intestinal Diseases
 - Brachyspira spp.
 - Anaerobic spirochetes requiring specialized selective media
 - Broadly classified as strongly beta-hemolytic and weakly betahemolytic
 - Infection is limited to the colon
 - Diarrhea of varying degrees and poor growth are observed
 - Diagnostics
 - Selective anaerobic culture
 - Speciation by PCR, MALDI-TOF, or sequencing
 - Direct PCRs
 - Histopathology + ISH or silver stains







- Large Intestinal Diseases
 - Brachyspira spp.
 - Swine Dysentery (SD)
 - Strongly beta-hemolytic species
 - B. hyodysenteriae
 - B. hampsonii
 - Clinical disease often triggered by stressors such as weaning and feed changes
 - Bloody and mucoid diarrhea
 - Varying the amount of dietary fiber can enhance or suppress clinical disease
 - Gross lesions
 - Mucohemorrhagic and fibrinous colitis



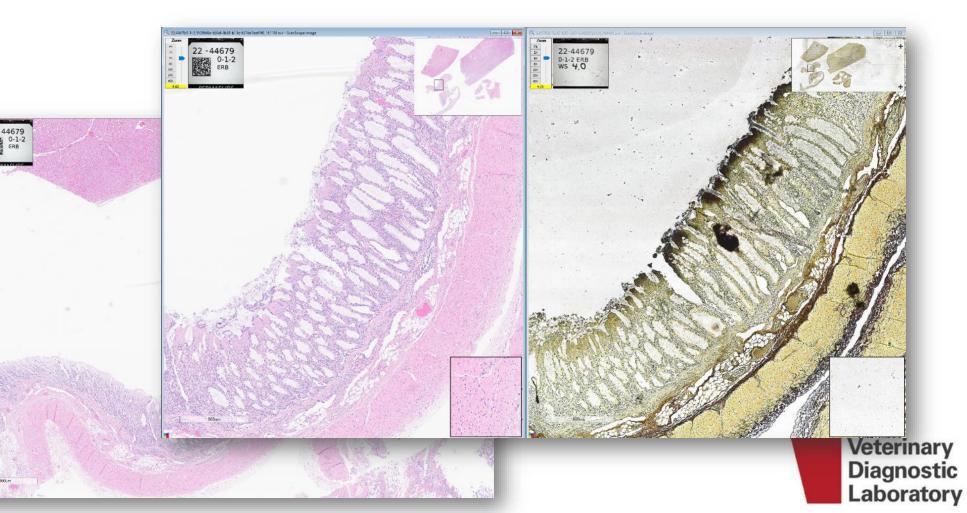




- Large Intestinal Diseases
 - Brachyspira spp.
 - Swine Dysentery (SD)

Histology (SD):

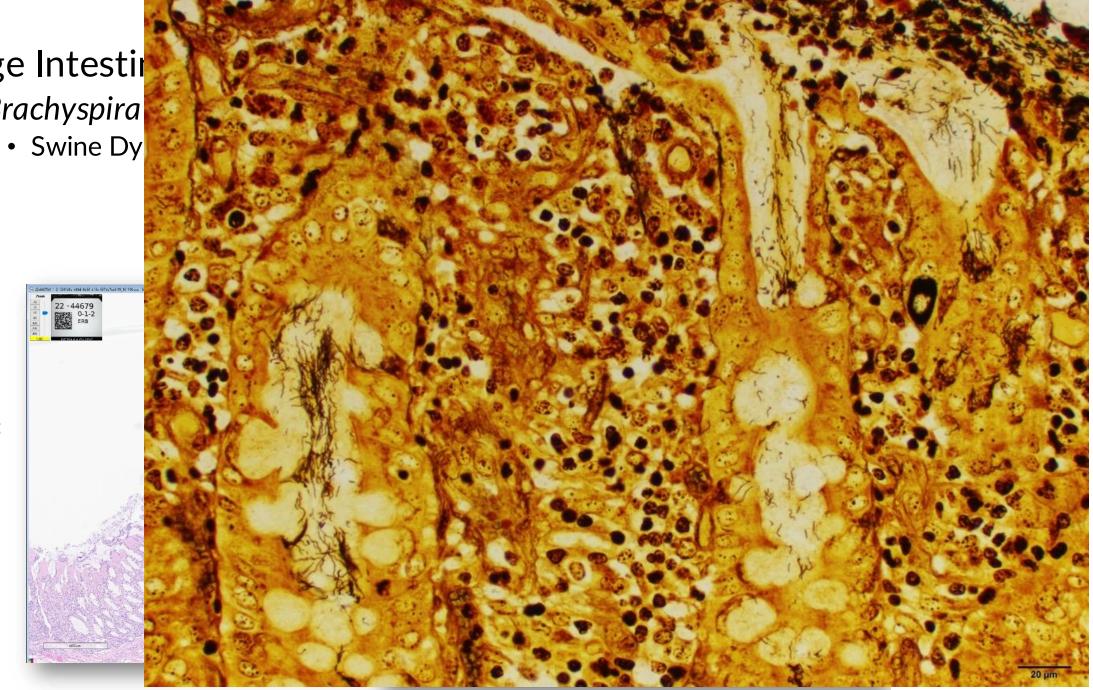
- Thickened mucosa
- Increased goblet cells
- Superficial hemorrhage
- Mucus efflux
- Mild to moderate mixed inflammation





- Large Intesti
 - Brachyspira

Histology (SD):



- Large Intestinal Diseases
 - Brachyspira spp.
 - Porcine Intestinal Spirochetosis (PIS)
 - Weakly beta-hemolytic
 - B. pilosicoli
 - Clinical disease often triggered by stressors such as weaning and feed changes
 - Mucoid diarrhea
 - Gross lesions
 - Mild to moderate mucoid colitis







- Large Intestinal Diseases
 - Brachyspira spp.
 - Porcine Intestinal Spirochetosis (PIS)

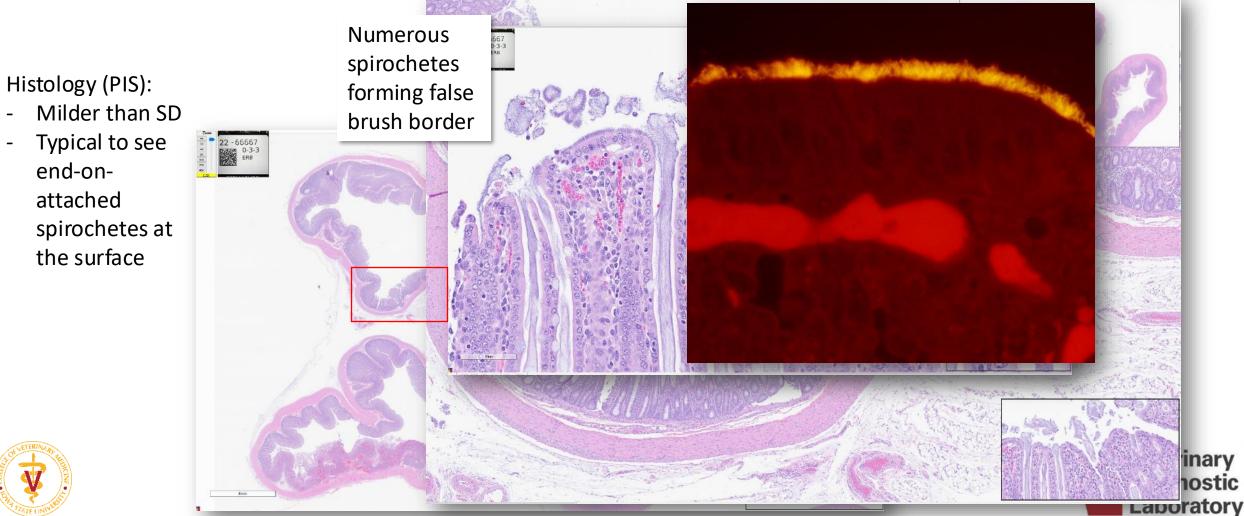
Histology (PIS):

- Milder than SD
- Typical to see end-onattached spirochetes at the surface



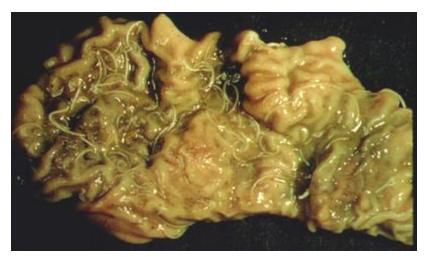


- Large Intestinal Diseases
 - Brachyspira spp.
 - Porcine Intestinal Spirochetosis (PIS)



- Large Intestinal Diseases
 - Trichuris suis
 - Diarrhea sometimes with blood and mucus
 - May resemble swine dysentery
 - Common in older, outdoor pigs



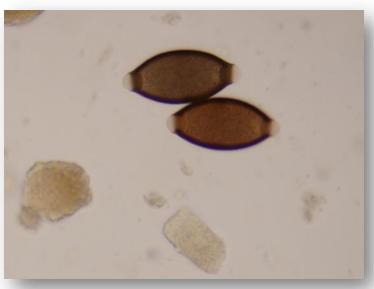


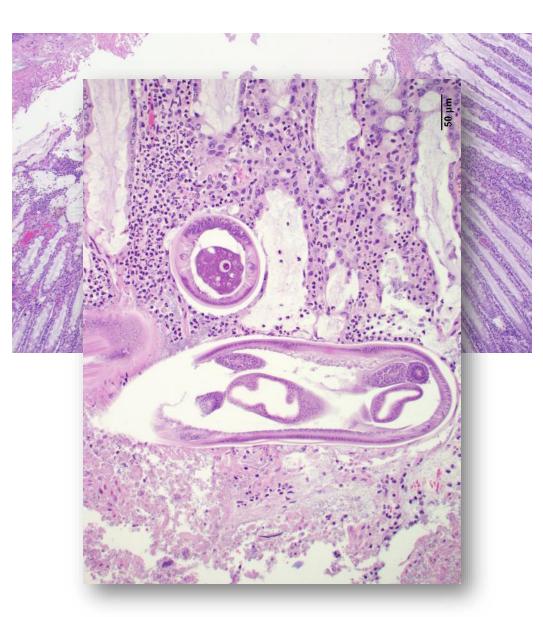


- Gross Lesions
 - Adult worms are ~2 inches long
 - Primarily in the cecum, may be in spiral colon



- Large Intestinal Diseases
 - Trichuris suis
 - Diagnostics:
 - Gross observation
 - Histopathology
 - Can occur with other diseases
 - Fecal flotation (long prepatent period)
 - Not useful for acute infections

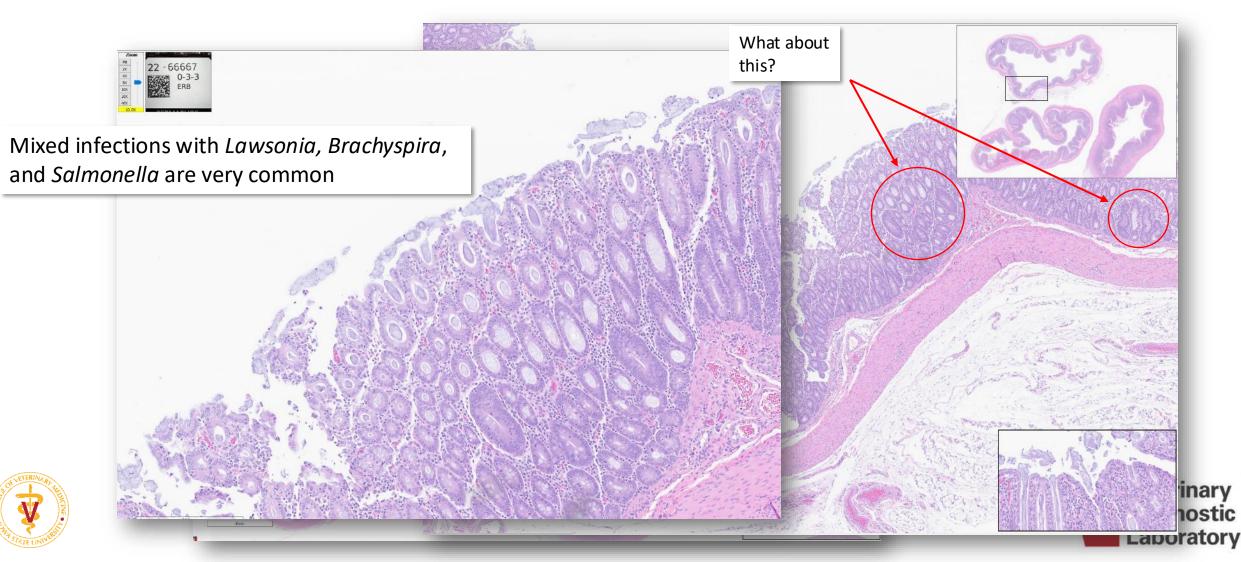




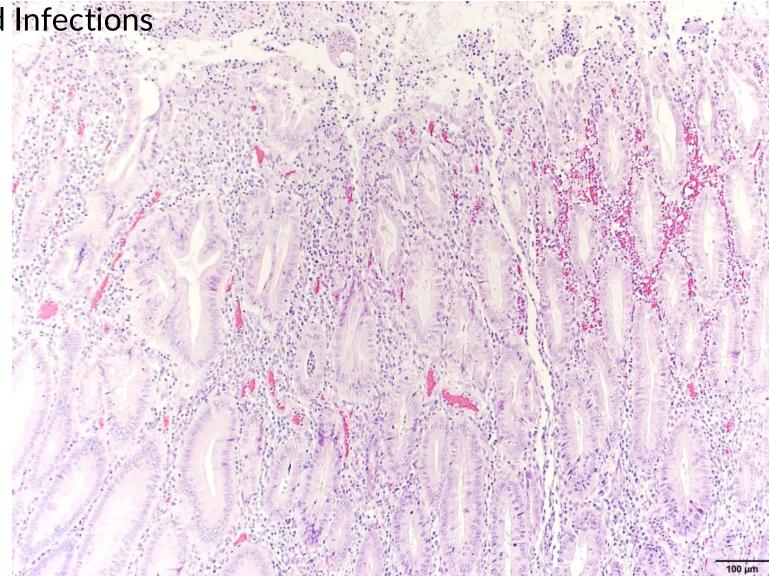




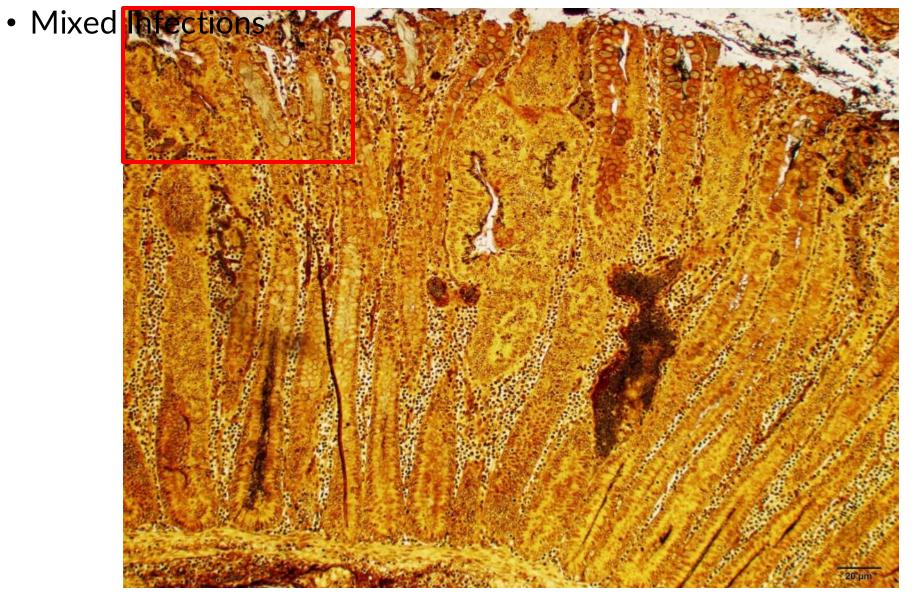
- Large Intestinal Diseases
 - Mixed Infections

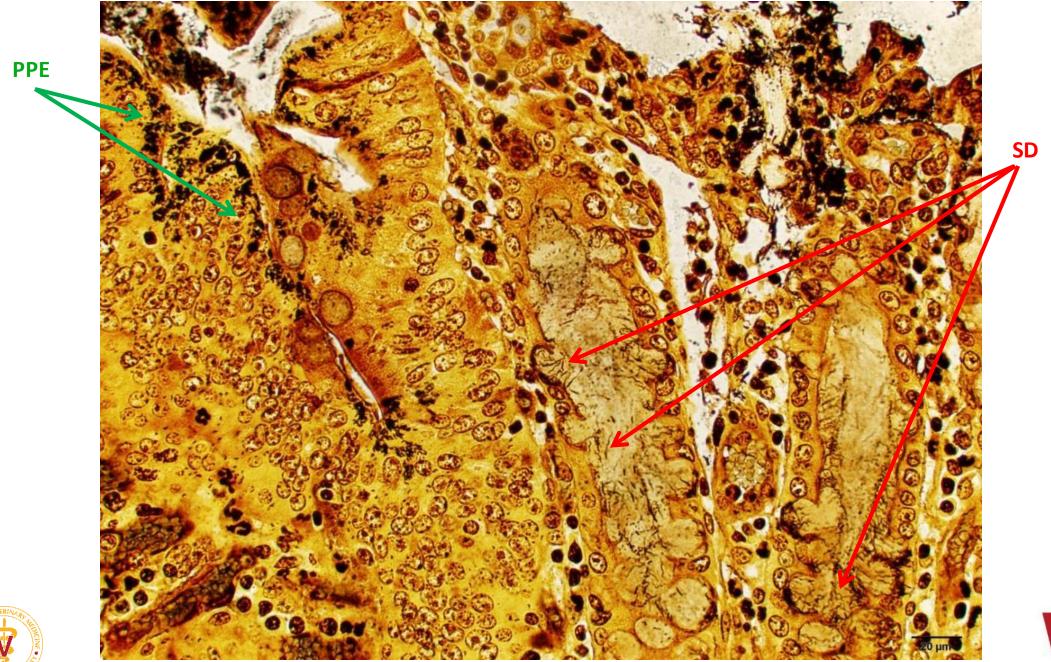


- Large Intestinal Diseases
 - Mixed Infections



• Large Intestinal Diseases







• Summary

- Health challenges in commercial pigs often present a diagnostic dilemma
 - Many potential pathogens are endemic in populations and disease expression is variable
 - Subclinical disease is common and can impact interpretation of treatment responses
- Very few lesions are pathognomonic
 - New agents/diseases/syndromes are being discovered every year in swine
 - In large part due to the increasing availability of NGS + ISH
 - PDCoV, PSaV, B. hampsonii, etc. are examples since 2012
 - Direct detection assays are available at ISU VDL for most of the agents discussed
- Consistent microscopic evaluation of a full set of tissues can help detect unexpected infectious diseases in swine; however, the focus is still on *lesions*

Veterinary Diagnostic

• Brain, heart, lung, liver, lymph node, kidney, spleen, small intestine, and colon



Questions?





Eric R. Burrough, DVM, PhD, DACVP | Professor Diagnostic Pathologist | Pathology Section Leader Iowa State University Veterinary Diagnostic Laboratory 1937 Christensen Drive | Ames, IA 50011 (515) 294-1950 | <u>burrough@iastate.edu</u> | @erburrough

