



New APL funded project

Understanding the national status of swine
dysentery & related diseases associated
with other *Brachyspira* spp.

Dr Rebecca Abraham

Dr Kittitat Lugsomya

Emeritus Prof. David Hampson*

Professor Sam Abraham



SWINE DYSENTERY (SD)

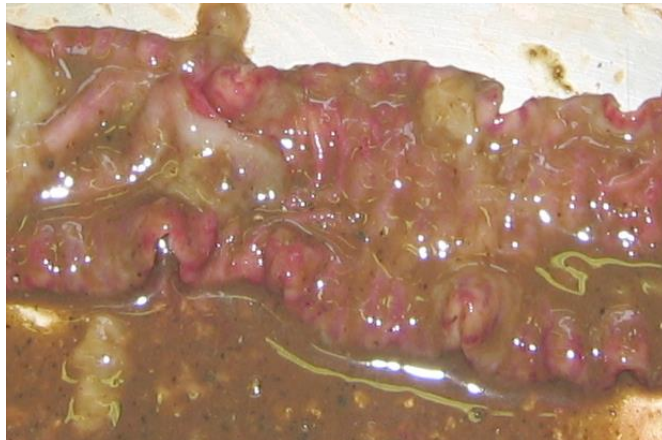
Classically = a severe mucohaemorrhagic colitis caused by infection with the intestinal spirochaete *Brachyspira hyodysenteriae*

- Mainly seen in growers and finishers



BUT

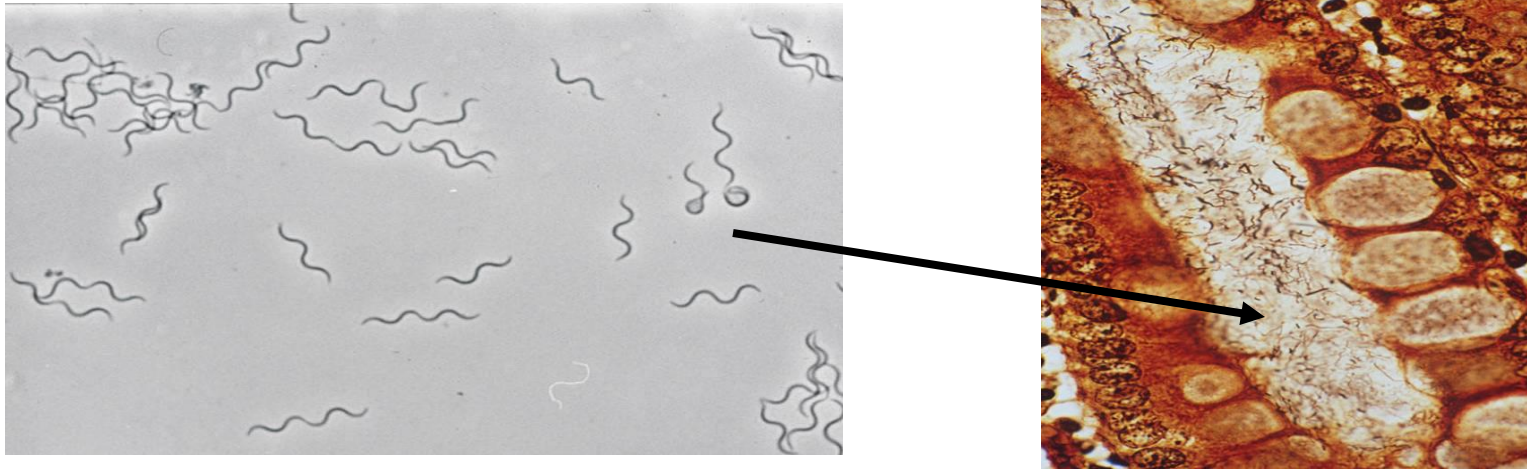
It is increasingly recognized that *B. hyodysenteriae* may be isolated from pigs in “healthy” herds



AND

two other *Brachyspira* spp. can cause SD

Brachyspira hyodysenteriae



An anaerobic intestinal spirochaete that colonises the large intestine of pigs, and occasionally other species

- Grows slowly (3+ days for growth to appear on plates)
- Requires selective agar to suppress other microorganisms
- Characteristically produces strong beta-haemolysis on blood agar

Haemolysis on blood agar

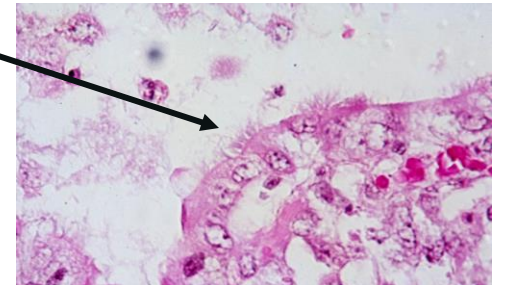
**Strong
haemolysis**



**Weak
haemolysis**

Brachyspira species

	β -haemolysis	Disease
<i>B. hyodysenteriae</i>	Strong	SD in Pigs; chickens
<i>B. hampsonii</i>	Strong	SD in Pigs
<i>B. suanatina</i>	Strong	SD in Pigs; mallards
<i>B. pilosicoli</i>	Weak	Intestinal spirochaetosis in Pigs, chickens, humans etc
<i>B. murdochii</i>	Weak	<u>Pigs?</u>
<i>B. innocens</i>	Weak	-
" <i>B. pulli</i> "	Weak	-
" <i>B. canis</i> "	Weak	-
<i>B. alvinipulli</i>	Weak	Chickens
<i>B. aalborgi</i>	Weak	Humans



The current study builds on our 2014-2016 Australian survey, funded by the Pork CRC

La T, Phillips ND and Hampson DJ (2016). An investigation into the etiological agents of swine dysentery in Australian pig herds. *PLoS ONE* 11(12): e0167424.

97 Australian herds of different reported health status sampled between June 2014 – June 2016

Samples (n=1103) mainly from healthy pigs obtained at the abattoir

- *B. hyodysenteriae* isolated from samples from 34 of the 97 herds (**35.1%**)
 - 25 herds reported as having a history of SD; 12 were culture positive (**48%**)
 - 48 herds reported as of uncertain health status; 8 were culture positive (**16.7%**)
 - 24 herds reported to be SD-free; 14 were culture positive (**58.3%**)

Findings from our 2014-2016 Australian survey: *B. hyodysenteriae* isolates

96 *B. hyodysenteriae* isolates recovered from the 34 infected herds (30 sequence types)

Multiple isolates recovered from 20 of the 34 herds

Nine of these 20 herds had between 2-4 different sequence types (STs) identified

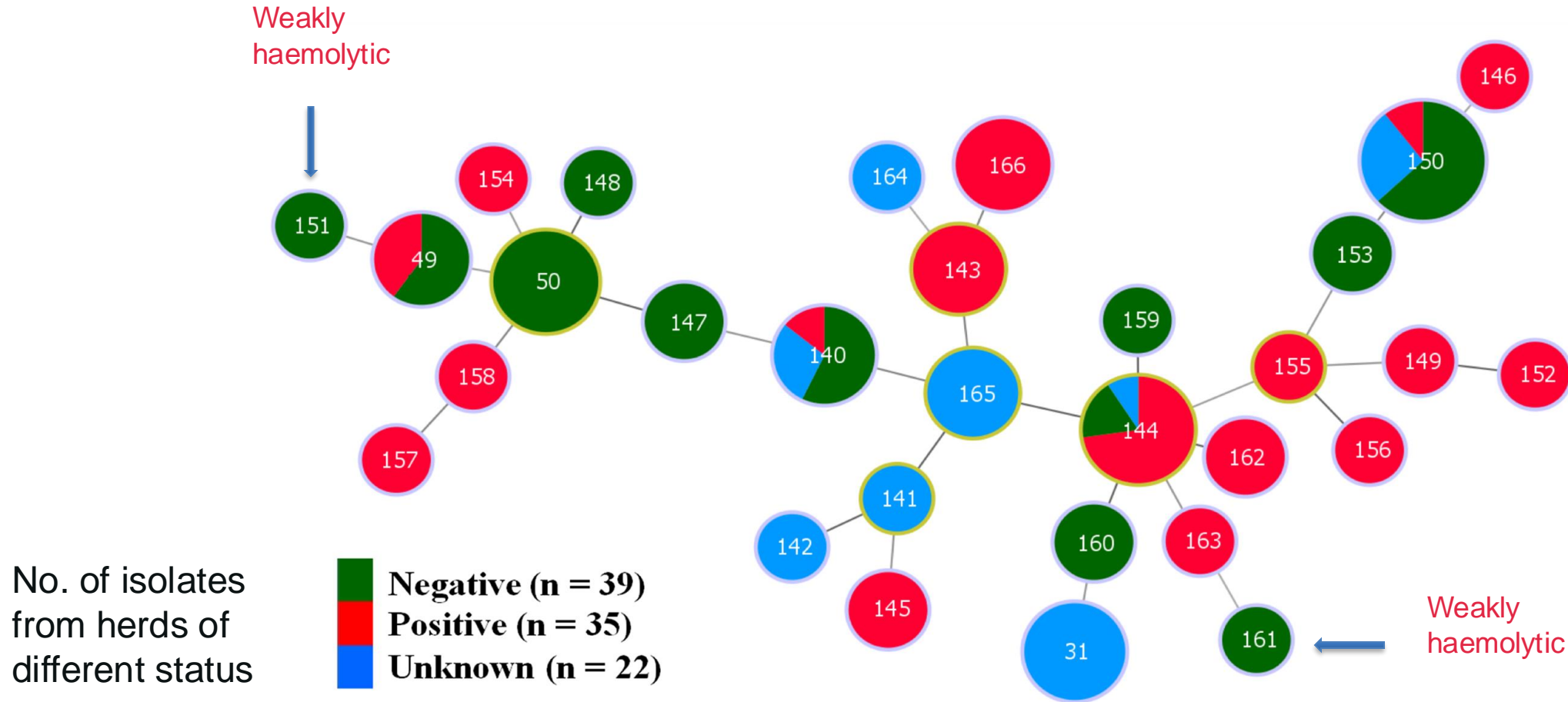
- ie multiple different strains present in these nine herd (including in four herds thought to be free of SD)

Four STs were shared by herds with different reported disease statuses

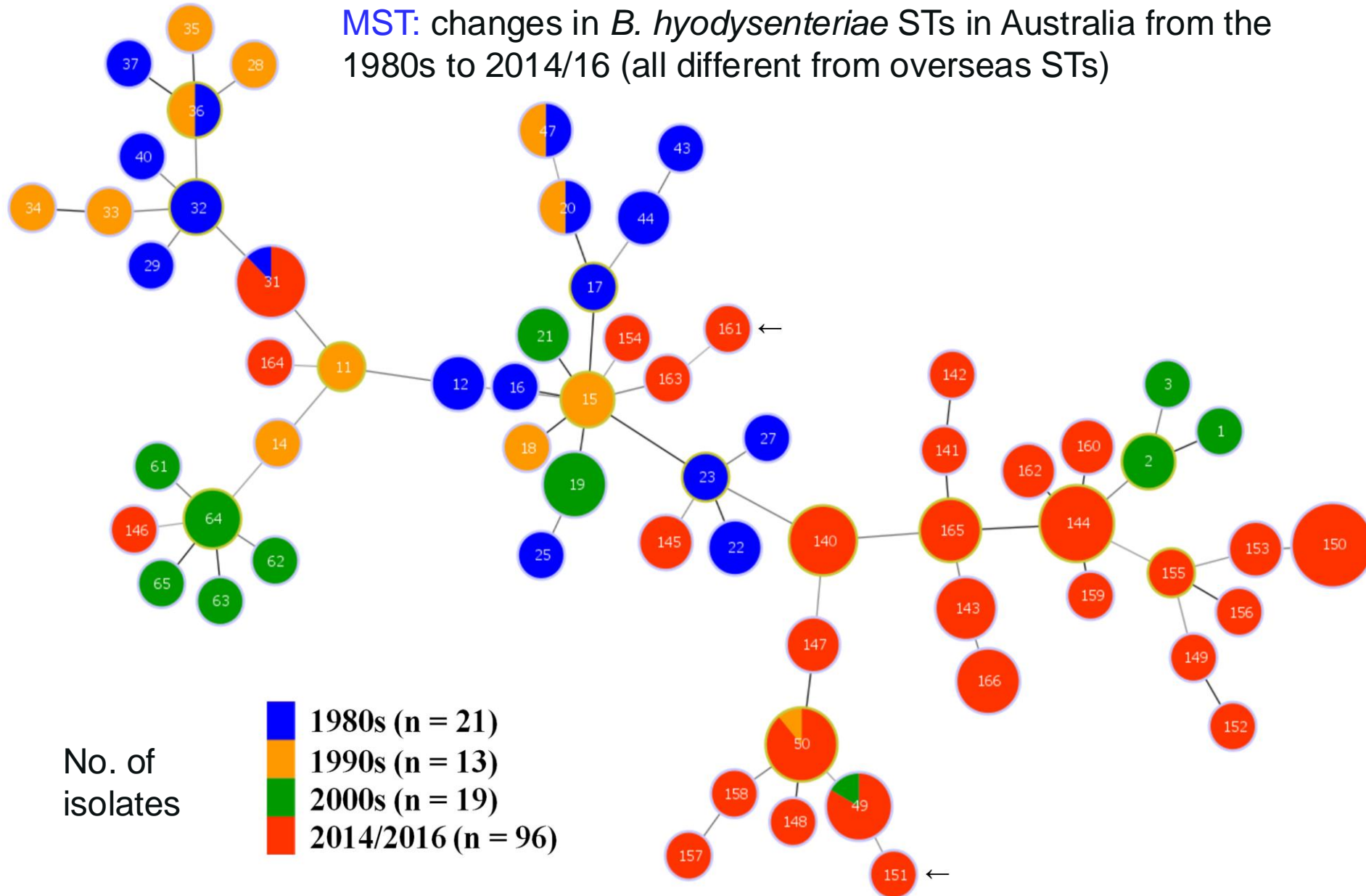
Two weakly haemolytic strains of *B. hyodysenteriae* identified

Minimum spanning tree (MST)

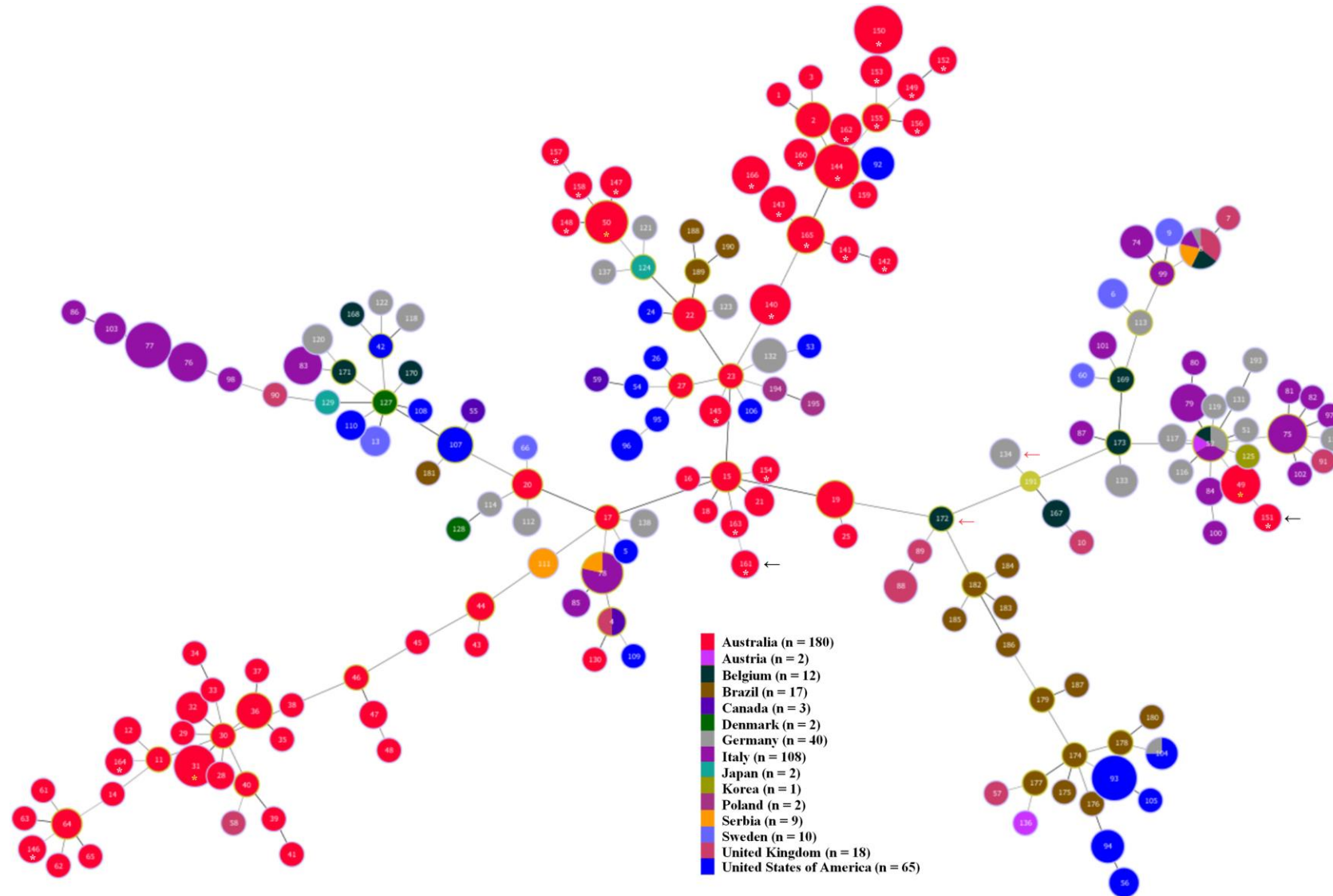
30 STs containing 96 *B. hyodysenteriae* isolates from 2014/16, from 34 herds with different reported health statuses



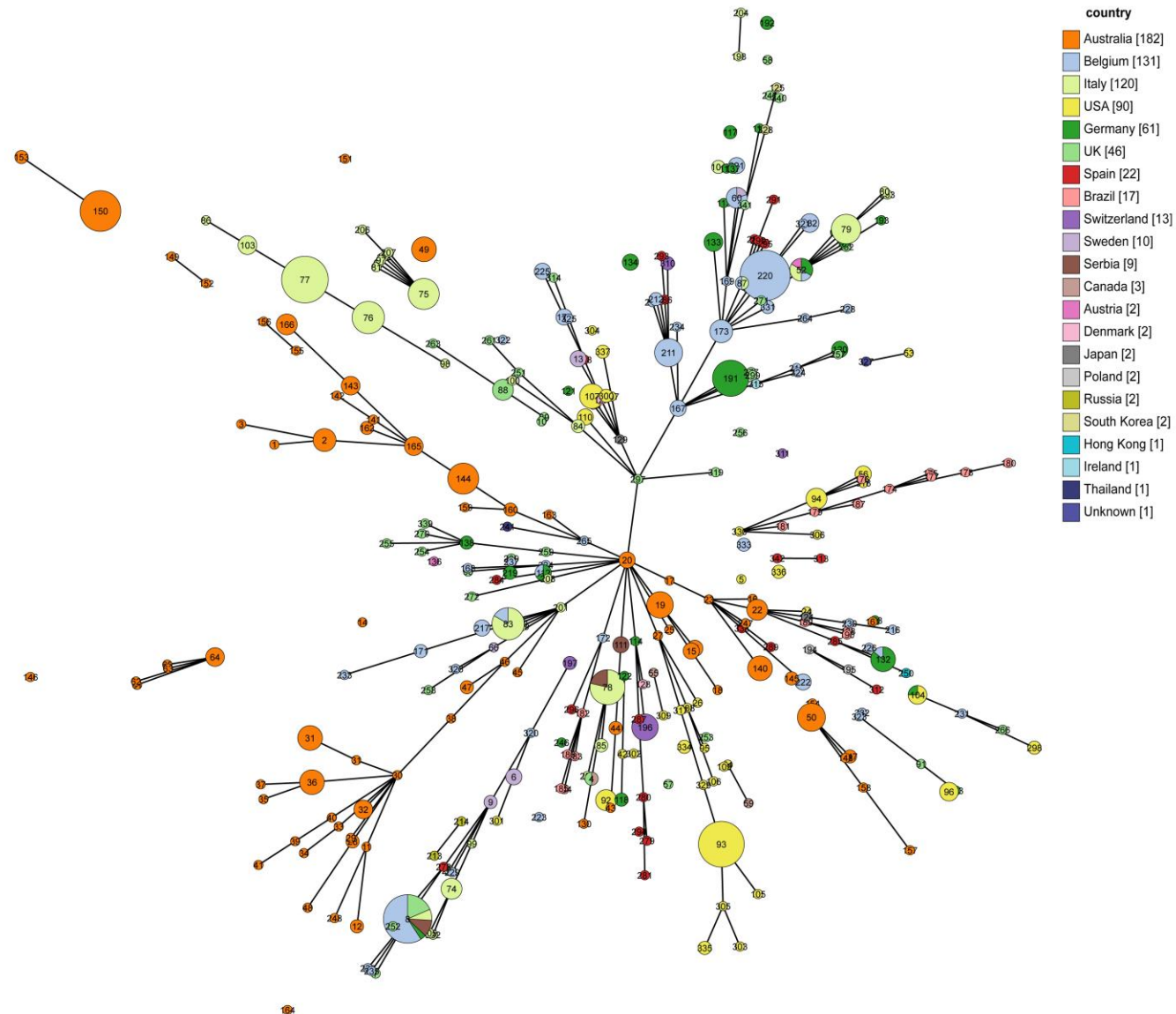
MST: changes in *B. hyodysenteriae* STs in Australia from the 1980s to 2014/16 (all different from overseas STs)



MST: International STs of *B. hyodysenteriae* in 2016



MST: International STs of *B. hyodysenteriae* in 2024



The current study builds on our 2014-2016 Australian survey, with the following aims

- To determine the prevalence of Australian herds currently colonized with *B. hyodysenteriae* and other *Brachyspira* spp.
- To characterize the genetic and phenotypic makeup of current strains, including their distribution and susceptibility to antimicrobials and potential virulence factors (using whole genomic sequencing)
- To compare current strains with those from previous decades (giving insights into the spread and evolution of the pathogen) and from herds of different health statuses
- To assist individual herds to determine their current *Brachyspira* status, and to investigate whether strains in infected herds have changed since the last survey

The current study

- Aiming for 100 herds of different health status
- Ideally 30 faecal/colonic samples from unmedicated growers/finishers per herd (doesn't need to be from diseased pigs but can accept samples from diseased pigs).
- *Brachyspira* spp. will be isolated, identified, and characterized by genomic sequencing and tested for antimicrobial susceptibility

Veterinarians and submitters – your participation is essential

- Please contact the lab before collecting/sending samples
- Record herd status (history of SD/uncertain/believed to be SD free)
- Answer survey questions about medication if possible
- The individual herd results will be returned to you (in confidence)
- An overview of results will be presented once the study is completed

Shipping

**Dr Rebecca Abraham
or Dr Kittitat Lugsomya (“Nick”)**

0413076742

0413347082

- Ship samples at **4°C overnight express** to:

Attn: AMRID

Loading Zone 5

Murdoch University

90 South Street

Murdoch WA 6150

Murdoch TNT account number 20118171

